



Seeding Cells in OptiCell®

Please always remember to use aseptic cell culture techniques when handling OptiCell units.

Materials

OptiCell units	1X PBS
OptiCell rack	50 ml centrifuge tubes
10 cc syringe or larger	Inverted microscope
Alcohol pads	Trypsin-EDTA
OptiCell Insertion tips	Sterile pipettes
Complete medium (compatible with cell line)	Sterile container for solution

Procedure

1. Remove OptiCell units from packaging inside a laminar flow hood. Place the OptiCell units into an OptiCell rack, port side facing out.
2. Aseptically place complete cell culture medium into an appropriate sterile container.
3. Attach a tip to a syringe and fill with 9 ml of complete growth medium.
4. Remove an OptiCell unit from the rack and insert tip into a port. Fill with 9 ml of medium. Without removing the tip, invert OptiCell and remove air. Repeat as necessary to fill each OptiCell.
5. Fill syringe with cell suspension. Add 1ml of cell suspension to each OptiCell. Gently shake the OptiCell a few times to disperse the cells.
6. Place OptiCell units horizontally into the rack and incubate rack at settings appropriate for cell line (*see user note for adherent cells*).

User Note for Adherent Cells:

1. To grow adherent cells on one surface (50 cm²), leave units in horizontal position.
2. To grow cells on both surfaces (100 cm²), place units in horizontal position as described. After partial attachment, invert the OptiCell units (180° rotation), and incubate for the remaining time in this orientation (*see Flipping Time*).

Important: *This partial attachment period (Flipping Time) will vary for every cell line.*

